

Technical Notification 0011

Evaluation of Sub-2 µm Media for Packing of Capillary Columns for Use in Proteomics (2)

In this independent study from EPFL-Proteomics Core Facility, multiple C18-RP-phases from Dr. Maisch HPLC GmbH, Germany were tested for identifying the sub-2 µm Bulk Media which shows the best efficiency for analyzing peptides from a HeLa cell lysate standard. The number of MS/MS identified peptides and proteins indicates the separation quality of the packed columns.

ID No.: 0011

Analytes: Thermo Fisher Scientific HeLa cell lysate
(P/N: 88328)

Thermo Fisher Scientific Cytochrom C Digest
(P/N:161089)

Column Dimension: 400 mm x 75 µm

Elution Type: Gradient

A: ACN/H₂O 2:98 + 0.1% Formic Acid

B: ACN/H₂O 90:10 + 0.1% Formic Acid

Long Gradient:

Time [min]	Flow Rate [µl/min]	%B
0	0.25	1
12	0.25	1
1105	0.25	24
125	0.25	38
126	0.25	90
129	0.25	90
130	0.25	1
150	Stop Run	0

Short Gradient:

Time [min]	Flow Rate [µl/min]	%B
0	0.25	1
12	0.25	1
30	0.25	35
35	0.25	90
38	0.25	90
40	0.25	1
55	0.25	1
55	Stop Run	0

Flow Rate:

250 nl/min

(due to increased backpressure: 200 nl/min for Exsil Mono)

Temperature:

50 °C

(due to increased backpressure: 55 °C for Exsil Mono)

Materials:

1 g Bulk Media

Reference:

ReproSil-Pur 120 C18-AQ, 1.9 µm
 (P/N: r119.aq.0001)

1.5 µm:

Exsil Pure 120 RP18, 1.5 µm
 (P/N: 5136767.0001)

ReproSil Saphir 100 C18, 1.5 µm
 (P/N: ra115.9e.0001)

Exsil Mono 100 C18, 1.5 µm
 (P/N: 5136782.0001)

Introduction:

Current gold standard packing material used at the EPFL-SV PCF Platform is the widely used and very efficient ReproSil-Pur 120 C18-AQ, 1.9 μm . As a permanent effort to improve analytical workflows, our group regularly performs series of comparative evaluations of different available packing materials. A set of Dr. Maisch products were tested to challenge the currently used setting including small particulate products (1.5 μm).

Chromatographic, Mass Spectrometric Settings and Analytes:

Columns were packed in-house using a standard bomb loader at 100 bar. Pulled tips from CoAnn Technologies (75 μm ID, 8 μm tip aperture: ICT36007508F-50) were used, and a packing length of 40 cm was consistently achieved for each column.

Analytes:

Thermo Fisher Scientific Hela cell lysate standard sample was injected in duplicates with different loading amounts (300 ng and 10 ng) on a pre-column prior to separation.

LC-MS Settings:

1. Chromatography: Thermo Fisher Scientific Vanquish Neo nano UHPLC
2. Mass Spectrometer: Thermo Fisher Scientific Tribrid Ascend

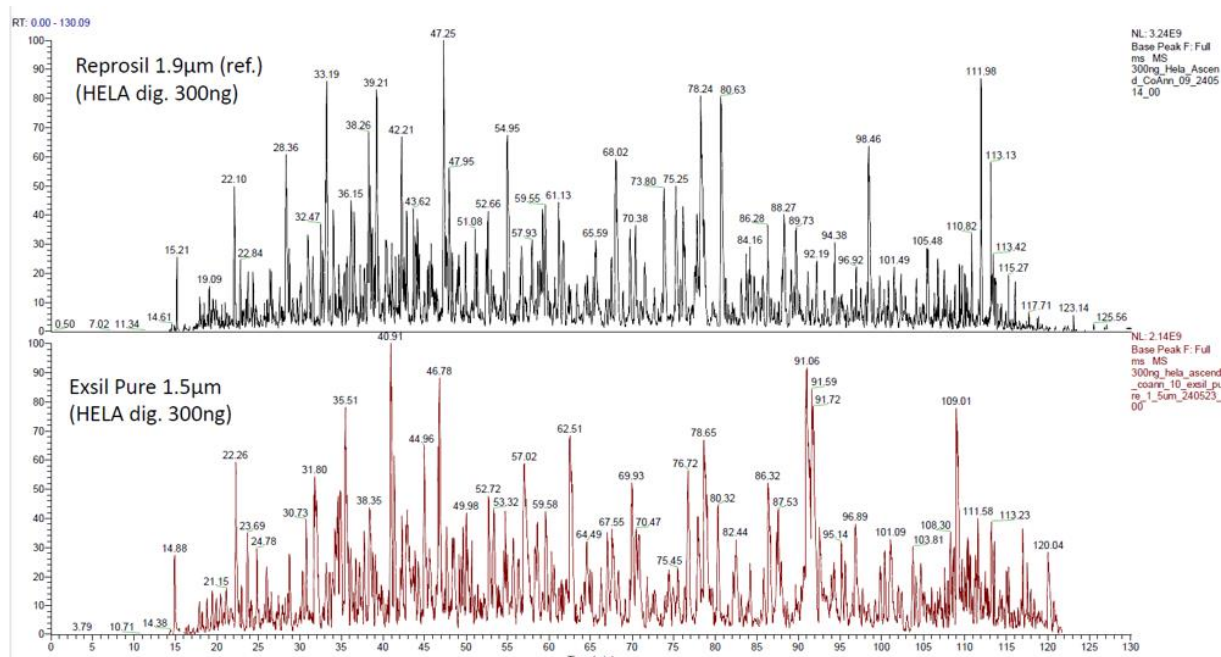
Data Processing:

MaxQuant search engine was used (ver.: 2.4.4.0). Basic settings:

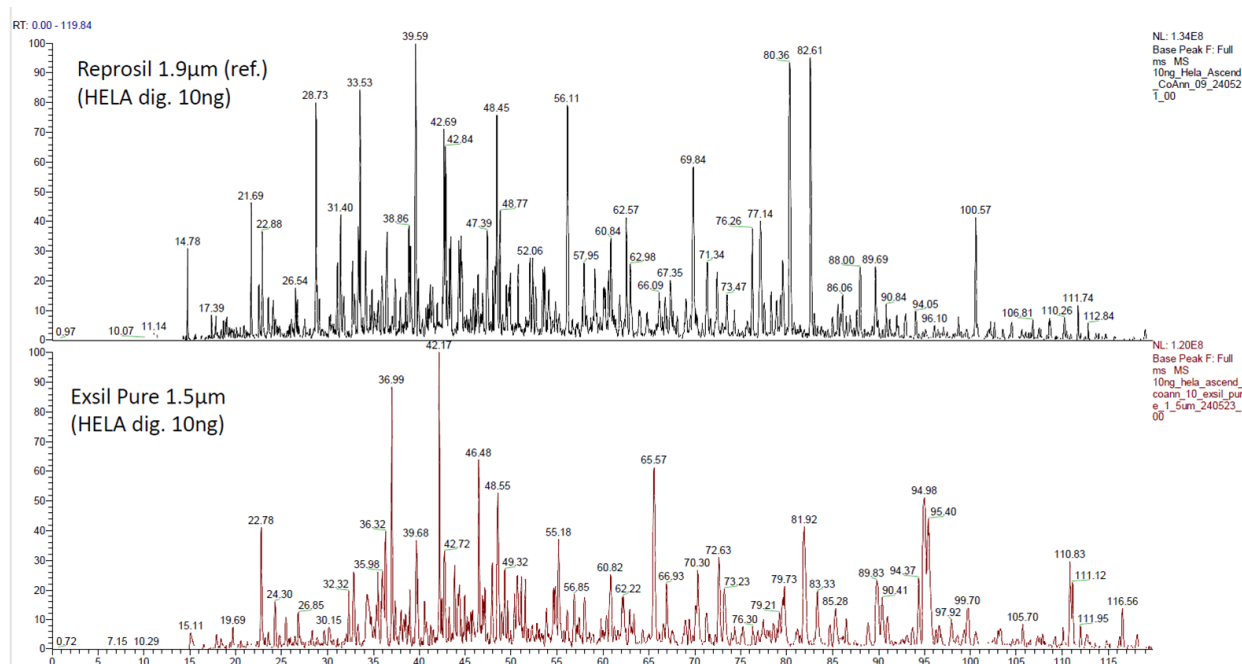
- Minimum of 2 peptides par proteins.
- 1% FDR threshold (PSMs, peptides and proteins).
- Variable modifications: oxidation (M), acetyl protein N term, phospho (STY).
- Fixed modifications: carbamidomethylation (C).
- Database: Uniprot_Human_74468Sequences_20190611.

Results:

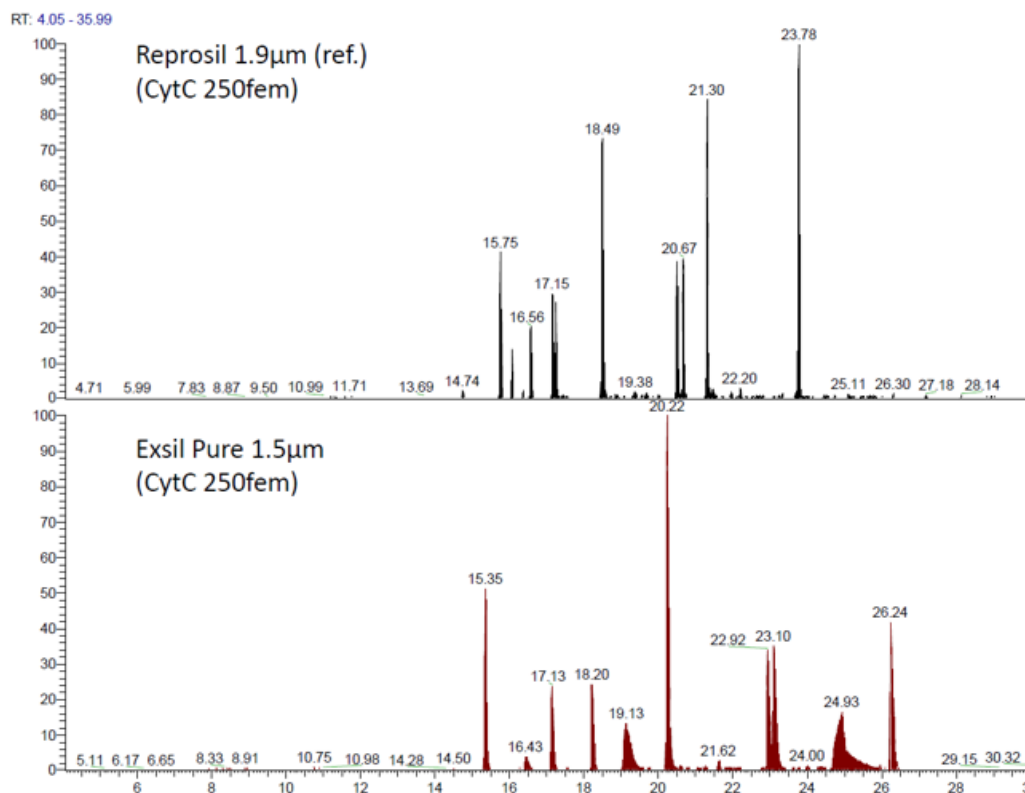
1a) Exsil Pure 120 RP18, 1.5 μ m (300 ng Injection HeLa cell lysate)



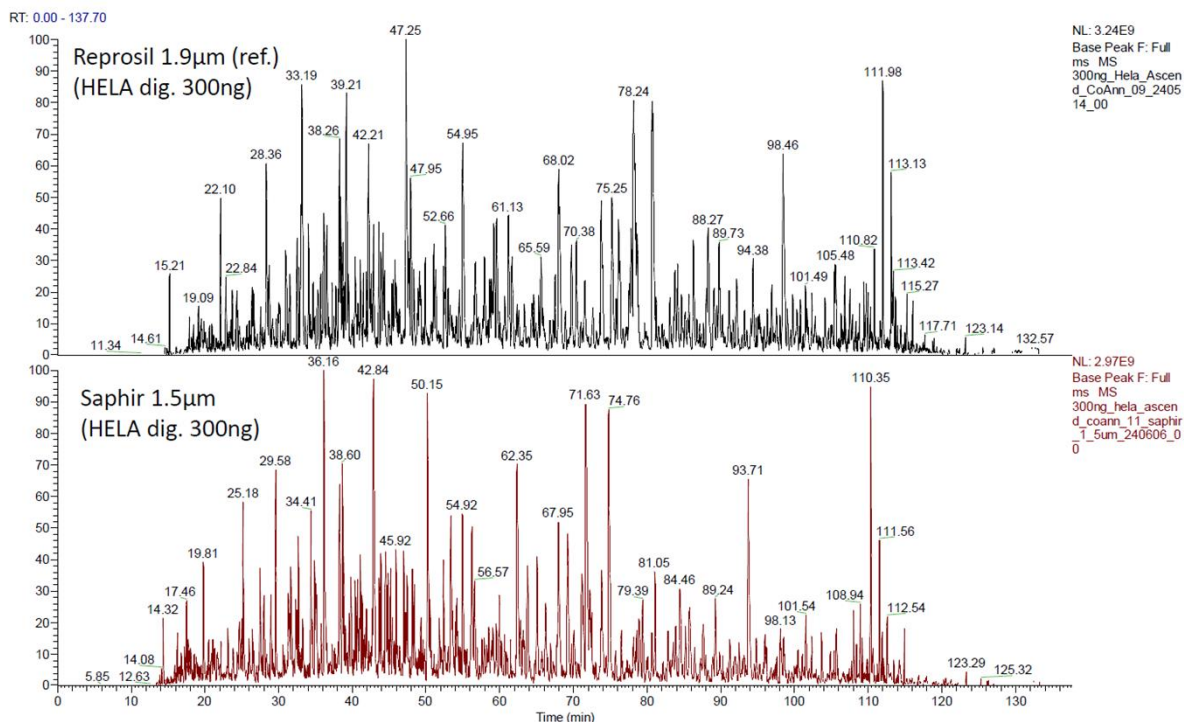
1b) Exsil Pure 120 RP18, 1.5 μ m (10 ng Injection HeLa cell lysate)



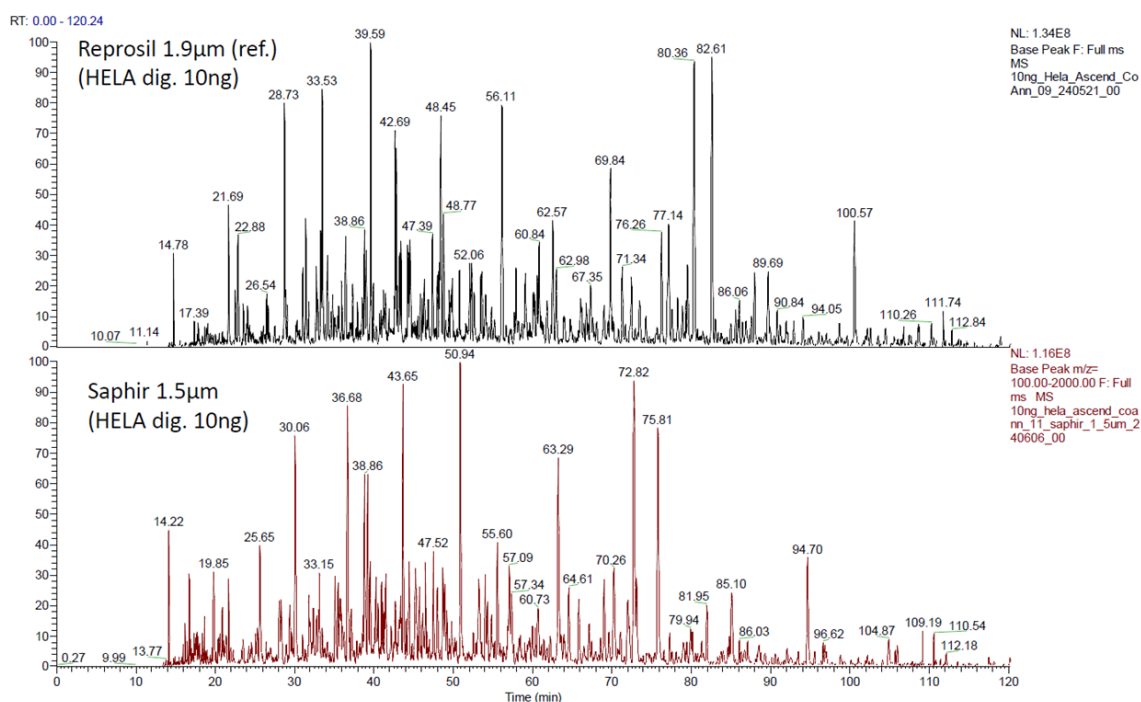
1c) Exsil Pure 120 RP18, 1.5 μ m (Cytochrom C Digest 250 fem (femtomole))



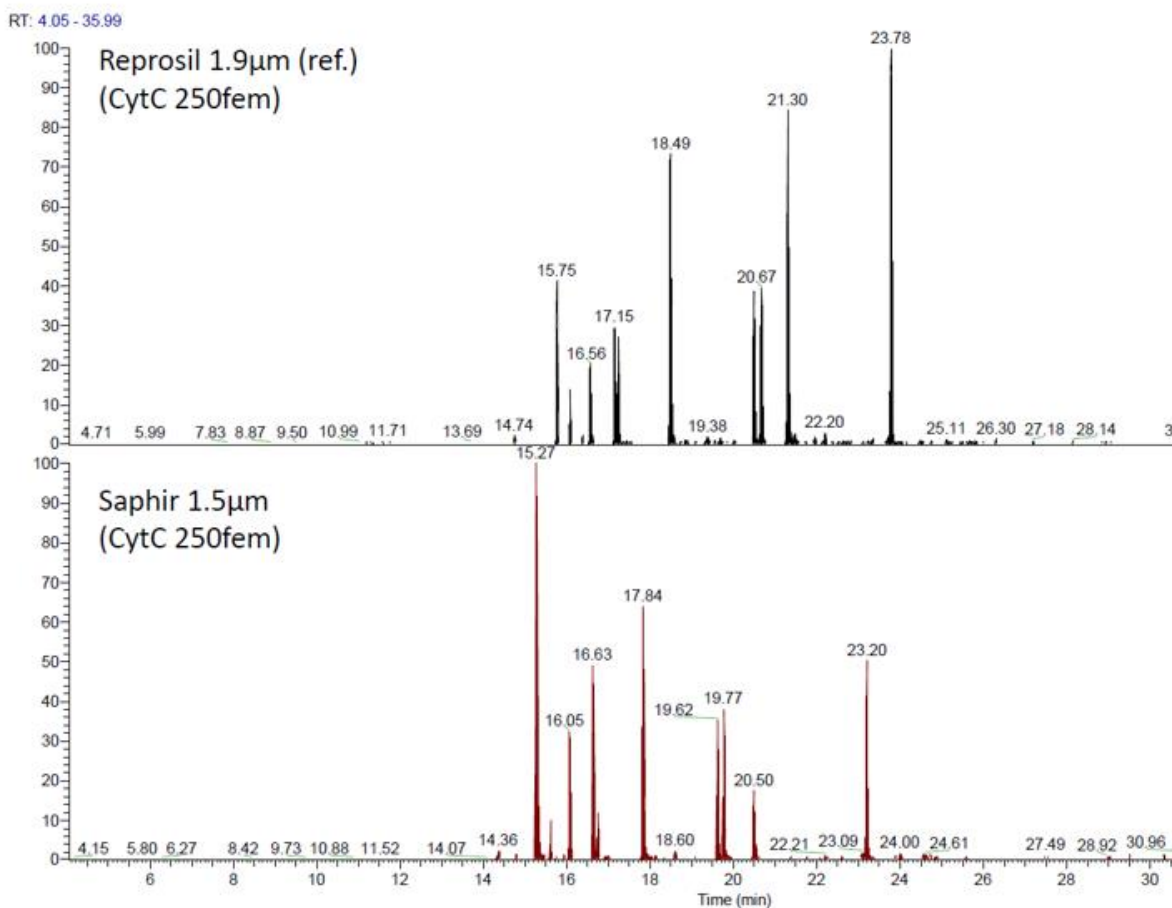
2a) ReproSil Saphir 100 C18, 1.5 µm (300 ng Injection HeLa cell lysate)



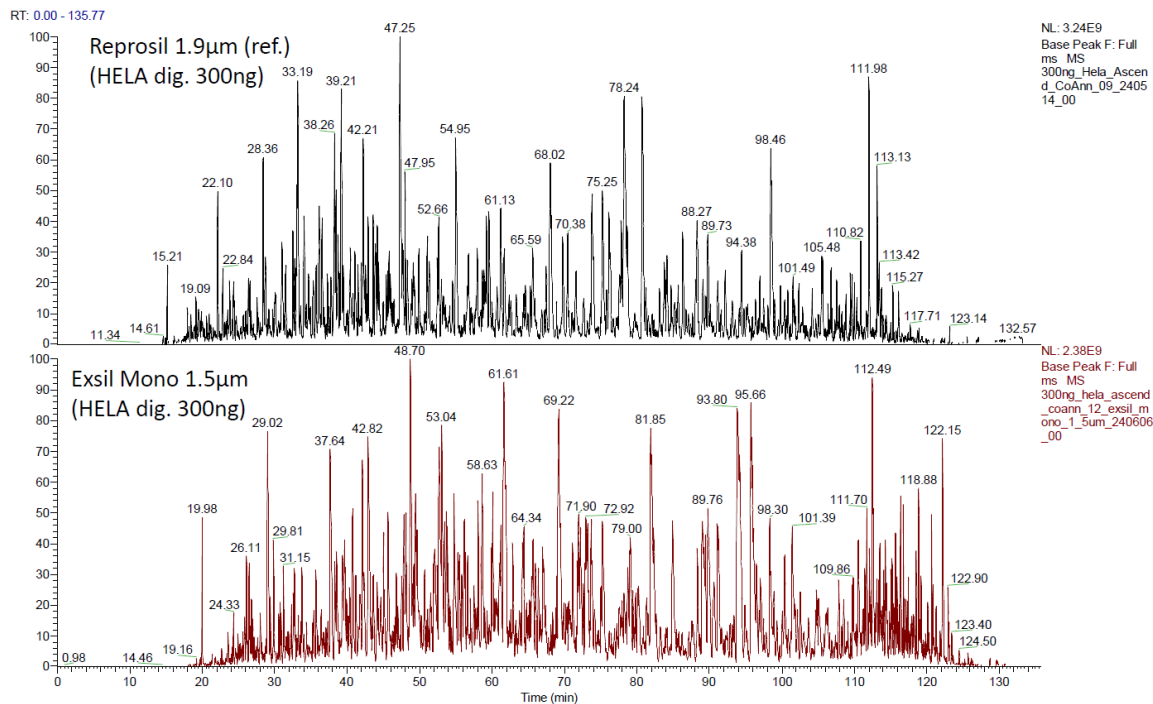
2b) ReproSil Saphir 100 C18, 1.5 µm (10 ng Injection HeLa cell lysate)



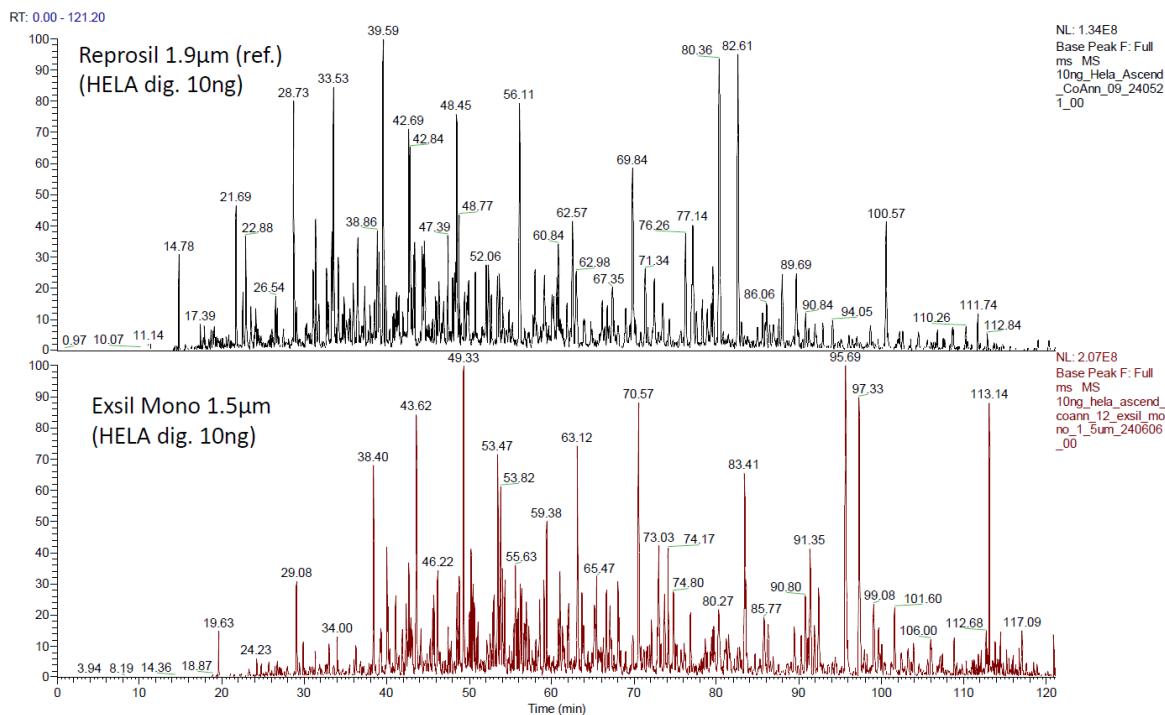
2c) Reprosil Saphir 100 C18, 1.5 μ m (Cytochrom C Digest 250 fem (femtomole))



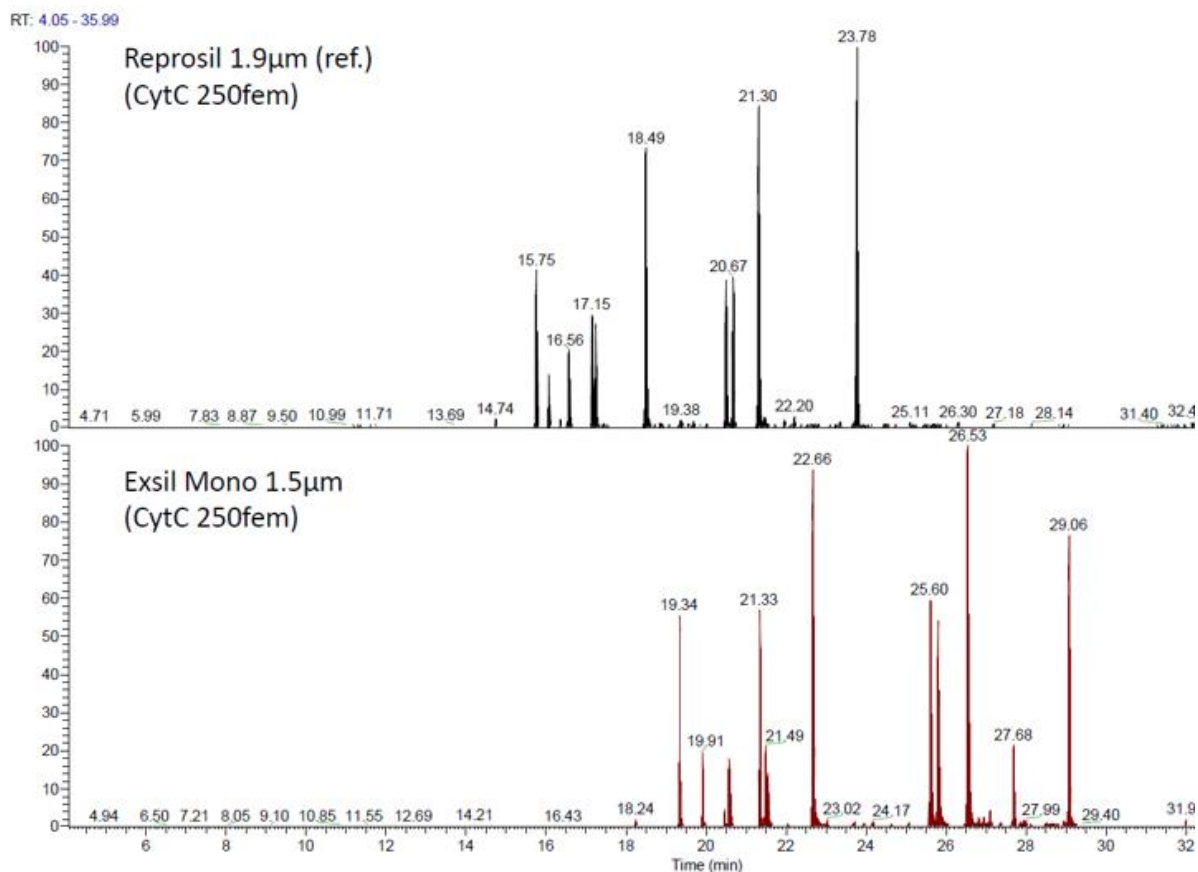
3a) Exsil Mono 100 C18, 1.5 µm (300 ng Injection HeLa cell lysate)



3b) Exsil Mono 100 C18, 1.5 µm (10 ng Injection HeLa cell lysate)



3c) Exsil Mono 100 C18, 1.5 μm (Cytochrom C Digest 250 fem (femtomole))



Identification Results:

Raw file	C18Material	MS	MS/MS	MS/MS submitted	MS/MS identified	MS/MS identified [%]	Peptide sequences identified	Peaks repeatedly sequenced [%]
300ng_Hela_Ascend_CoAnn_09_240514_00	Reprosil Pur AQ 1.9um	10'455	180'704	204'108	67'947	33	46'782	12
300ng_Hela_Ascend_CoAnn_09_240514_01	Reprosil Pur AQ 1.9um	10'447	180'444	204'020	68'705	34	46'856	12
300ng_Hela_Ascend_CoAnn_10_Exsil_Pure_1_Sum_240523_00	Exsil Pure 1.5um	10'657	189'476	226'000	67'128	30	42'046	18
300ng_Hela_Ascend_CoAnn_10_Exsil_Pure_1_Sum_240523_01	Exsil Pure 1.5um	10'632	188'588	223'106	66'834	30	42'334	17
300ng_Hela_Ascend_CoAnn_11_Saphir_1_Sum_240606_00	Saphir 1.5um	10'423	177'288	198'863	71'311	36	42'668	16
300ng_Hela_Ascend_CoAnn_11_Saphir_1_Sum_240606_01	Saphir 1.5um	10'435	177'630	199'193	70'649	35	43'104	16
300ng_Hela_Ascend_CoAnn_12_Exsil_Mono_1_Sum_2406012_00	Exsil Mono 1.5um	10'510	187'091	217'973	70'998	33	49'631	11
300ng_Hela_Ascend_CoAnn_12_Exsil_Mono_1_Sum_2406012_01	Exsil Mono 1.5um	10'510	186'389	217'065	70'605	33	49'819	11

Conclusion:

It has to be first mentioned that a completely fair comparison requires an entire control of every step of the process to avoid introduction of external variability (sample batch and dilutions, MS performance). We tried to maintain source of variations as low as possible keeping in mind an error tolerance would be considered at the end of the comparison process. Moreover, a single column was packed per chromatographic phase tested and eventual packing process variability has also to be considered. In our hands and over a couple of years of packing experience, this last variability source is rather limited using ReproSil-Pur 120 C18-AQ, 1.9 µm material (below 7%).

It's important to recognize that each chromatographic phase is better suited to specific applications or separation conditions (mobile solvents, additives, loading amounts, flow rates, etc.). Therefore, identifying a universal packing material might be challenging, and a compromise may be necessary.

In our hands, Exsil Mono material emerged as one of the best media tested. However, it required a longer packing time and exhibited higher backpressure compared to other evaluated C18 media.

The typical backpressure for the tested capillary columns ranges from 5000-6500 psi (345-448 bar). Exsil Mono material's backpressure was significantly higher, around 8000 psi. Due to the higher backpressure observed with Exsil Mono, some adaptations were necessary. We used a lower flow rate (200 nl/min instead of 250 nl/min) and applied a higher column heating temperature to ensure safe testing.

As a general trend, all tested materials showed good performance compared to the control column, especially when analyzing higher sample amounts (300 ng).

These comparisons, performed under the described conditions, highlighted Exsil Mono 100 C18, 1.5 μ m packing material as a very competitive alternative option to our gold standard material (ReproSil-Pur 120 C18-AQ, 1.9 μ m).

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Data courtesy of Dr. Diego Chiappe, Romain Hamelin EPFL-Proteomics Core Facility, Lausanne.

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