

# **Multi-Selectivity Stationary Phases**

## **By Dr. Maisch HPLC GmbH**

*A State-of-the-Art Solution for HPLC, UPLC and  
preparative LC*

## Selectivity in HPLC

Selectivity are mainly driven by the interactions of sample molecule with the surface of the stationary phase supported by the mobile Phase

# Possible Interactions in RP-HPLC

## Phase / Sample interactions:

- |  |           |
|--|-----------|
| • Lipophilic   | Non-polar |
| • Dipole – induced dipole<br>• Dipole – Dipole<br>• Hydrogen bonding<br>• $\pi$ complex bonding<br>• cation exchange | polar     |

## Multi-Selectivity in HPLC

Multi – Selectivity means that the Stationary Phase offers different possibilities for interactions to support and increase selectivity

## Multi-Selectivity

### Lipophilic – Polar Phases

- Not endcapped C18 / C8 / Phenyl
- AQ / HE
- EPS
- Cyano-Propyl
- Amino-Propyl
- Diol

## Multi-Selectivity

### Lipophilic – $\pi$ -electron (Phenyl) Phases

- Propyl-Phenyl
- Hexyl-Phenyl
- Biphenyl
- Diphenyl
- C18-Phenyl
- C18-PFP
- PFP

## Multi-Selectivity

### Lipophilic – Ion exchange Phases

#### Mixed Mode Phases

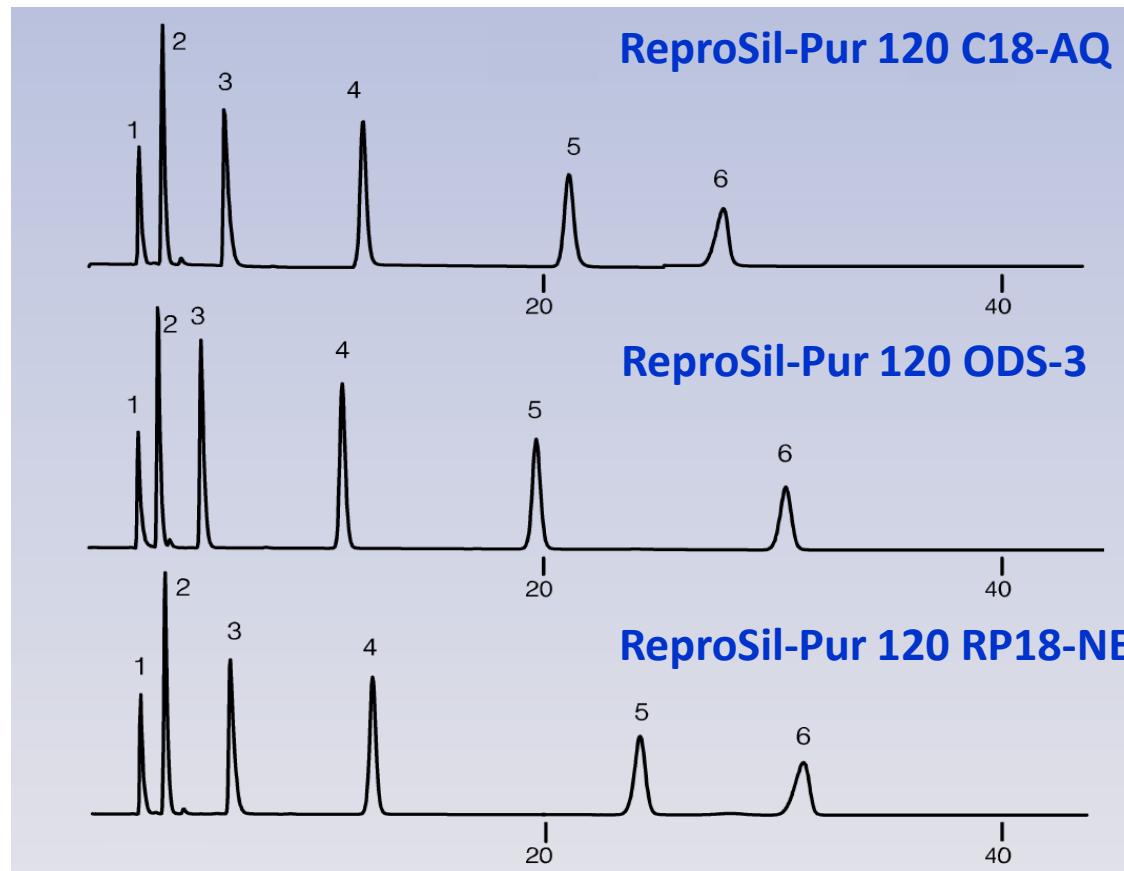
- C18 / SAX
- C8 / SAX
- C18 / SCX
- C8 / SCX
- Phenyl-SCX
- Propyl-SO<sub>3</sub>H
- Propyl-Amino

# Multi-Selectivity

## Hilic - Phases

- Si
- EPS
- Cyano-Propyl
- Amino-Propyl
- Diol
- Phenyl-SCX
- SCX
- SAX
- Zwitter Ionic
- ... special polar groups

# Lipophilic – Polar Phases

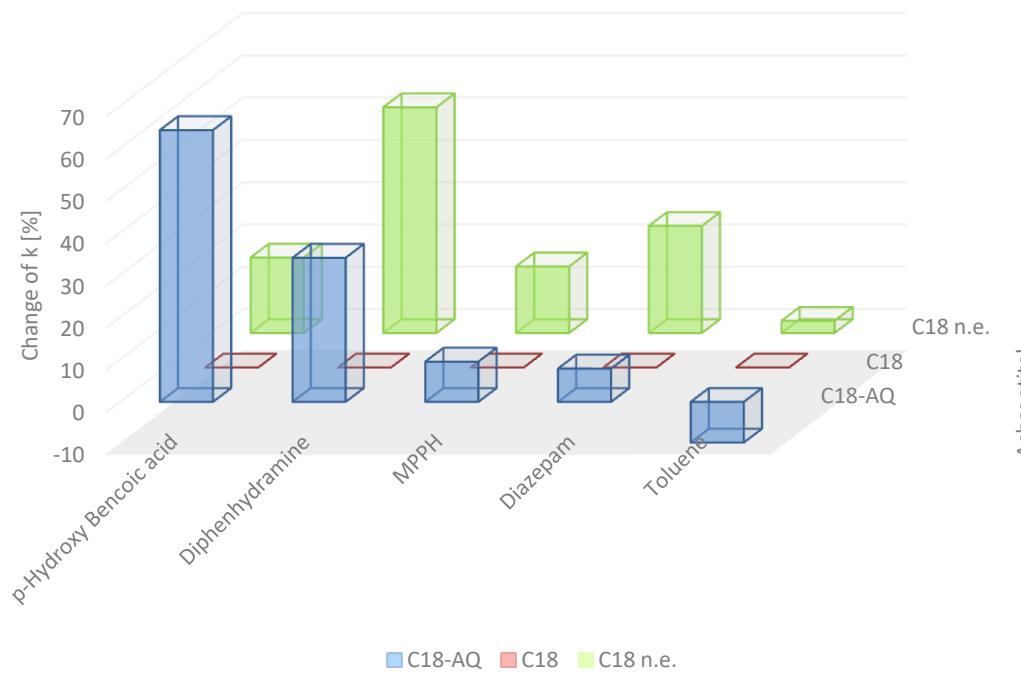


- 1) uracil
- 2) 4-hydroxybenzoic acid
- 3) diphenylhydramine
- 4) 5-(p-methylphenyl)-5-phenylhydantoin
- 5) diazepam
- 6) toluene

Flow rate: 1.0 ml/min  
Eluent: Na-phosphate buffer, 50 mM, pH 2.3 /  
acetonitrile = 58 /42  
Detection (UV):230 nm  
NovoGrom 250 mm x 4.0 mm id.

# Lipophilic – Polar Phases

Selektivity differences of C18 Phases



# Selectivity in HPLC Stationary Phases

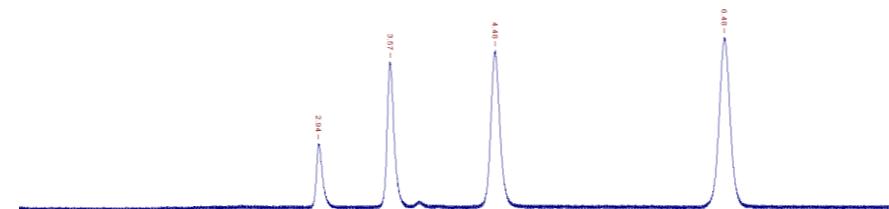
Phase	%C	Description
Reprospher 100 C30-DE	20%	double endc
Reprospher 100 C18	16%	endc.
Reprospher 100 C18-DE	16%	double endc
Reprospher 100 C18-Aqua	12%	pol.Gr.+ endc.
Reprospher 100 C18-Aqua-DE	12%	pol.Gr.+ double endc.
Reprospher 100 C18-NE	15%	not endc
Reprospher 100 C18-TDE	20%	polymer-C18-de
Reprospher 100 C18-TN	17%	polymer-C18, not endc.
Reprospher 100 C18-Phenyl		C18+Phenyl-Groups, endc.
Reprospher 100 C18/WCX		C18+carboxylic acids
Reprospher 100 C12	8%	endc.
Reprospher 100 C8-Aqua	8%	pol.Gr.+ endc
Reprospher 100 C8-Aqua-DE	8%	pol.Gr.+ d.endc.
Reprospher 100 C8	10%	endc.
Reprospher 100 C8-DE	10%	double endc.
Reprospher 100 C8-NE	9%	not endc.
Reprospher 100 C8-TN		polymer-C8, not endc.
Reprospher 100 Phenyl	9%	double endc.
Reprospher 100 Phenyl	9%	double endc.
Reprospher 100 Phenyl-NE	12%	not endc.
Reprospher 100 Phenyl-Hexyl	13%	not endc.
Reprospher 100 Phenyl-Hexyl-e	13%	endc.
Reprospher 100 Diphenyl (Phe-Si-Phe)		double endc.
Reprospher 100 Biphenyl (Si-Phe-Phe)		double endc.
Reprospher 100 PFP (Pentafluorophenyl)		double endc.

# Selectivity in HPLC Stationary Phases - 2

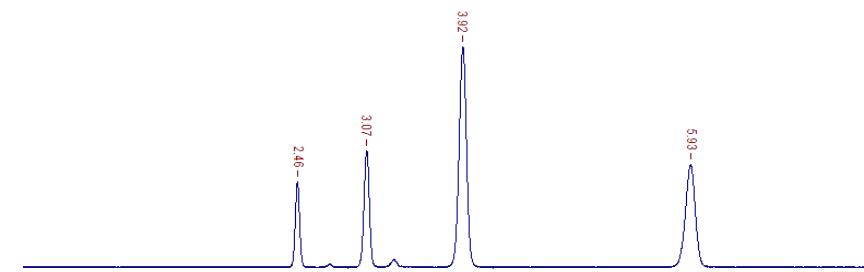
Reprospher 100 C6-TDE	8%	trifunct. endc.
Reprospher 100 C4-Aqua	6%	pol.Gr.+ endc.
Reprospher 100 C4	6%	endc.
Reprospher 100 C4-DE	7%	double endc.
Reprospher 100 CN	7%	
Reprospher 100 CN-DE	7%	double endc.
Reprospher 100 Diol-DE	7%	double endc.
Reprospher 100 Diol	7%	
Reprospher 100 2-EP		2-Ethylpyridin
Reprospher 100 4-EP		4-Ethylpyridin
Reprospher 100 NH2-DE	4%	endc.
Reprospher 100 NH2	4%	not endc.
Reprospher 100 DNH	5%	Diamin
Reprospher 100 PEI, Polyethylenimin		WAX for peptides / Nucleotide
Reprospher 100 Si		
Reprospher HILIC-P		HILIC for Peptides
Reprospher HILIC-A		HILIC for Acids
Reprospher HILIC-ARG		(with Arginin-Group)
Reprospher Acidosil-S		SO3H-Silica
Reprospher Acidosil-C		COOH-Silica

# Selectivity in HPLC D4 Vergleich C18, Phenyl, C18-Phenyl

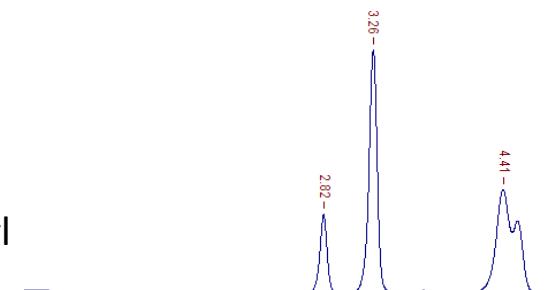
D4 Reprospher 100 C18



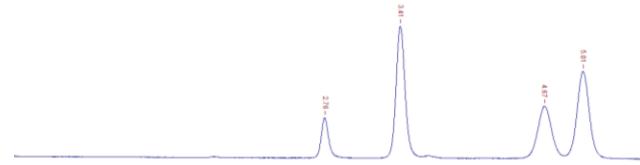
D4 Reprospher 100 C18-DE



D4 Reprospher 100 C18-Phenyl

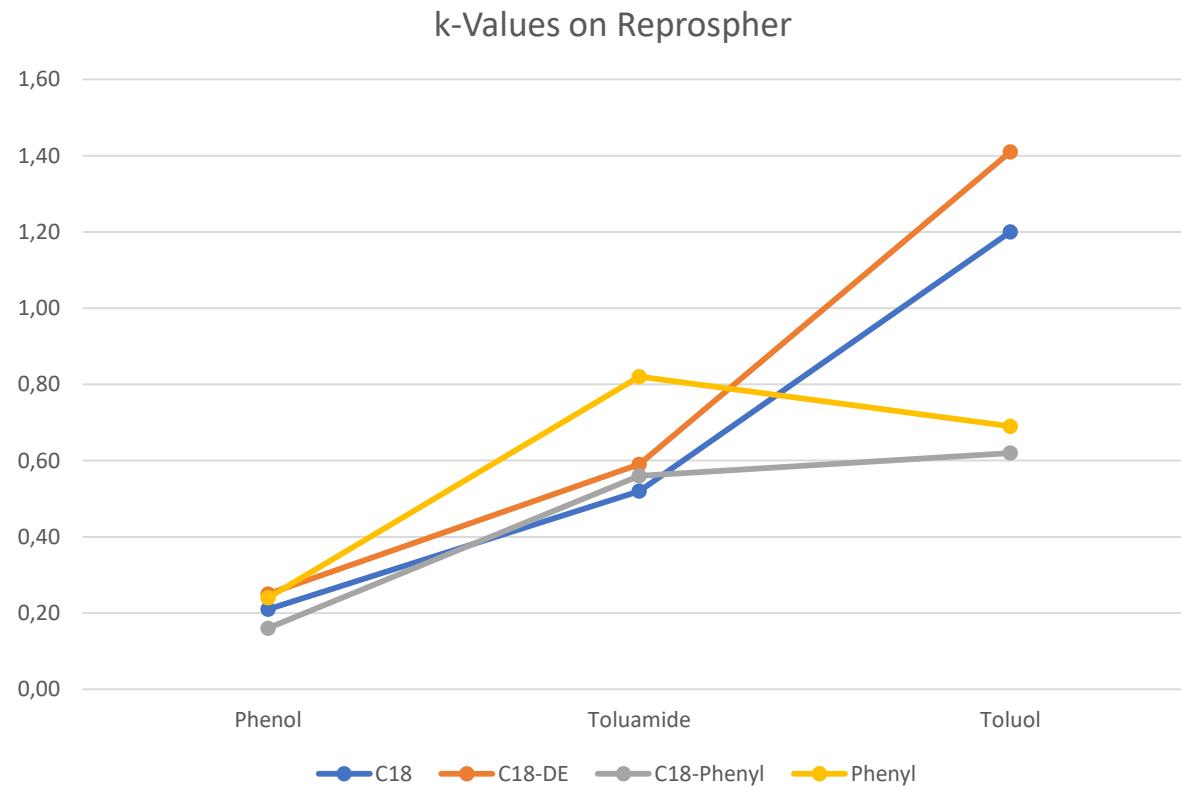


D4 Reprospher 100 Phenyl

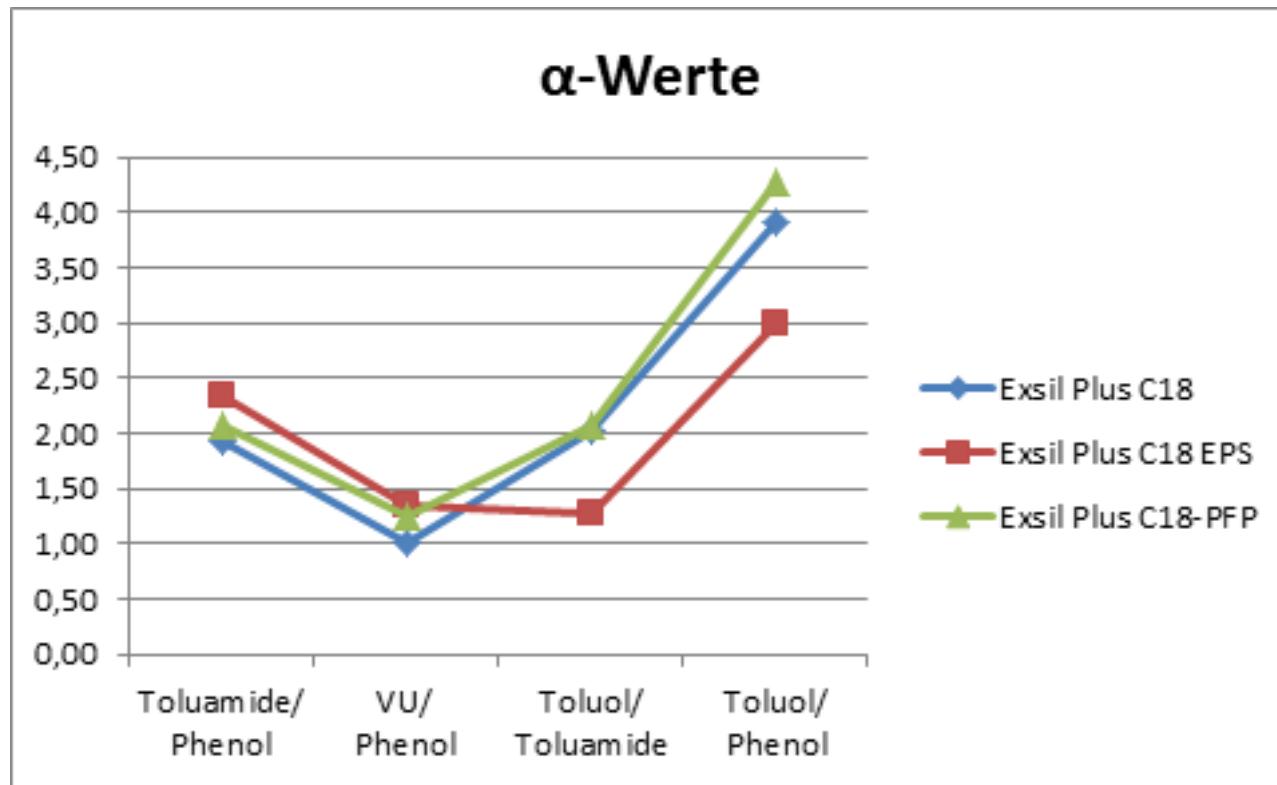


# Selectivity in HPLC

D4-Test: C18, Phenyl, C18-Phenyl



# Selectivity in HPLC

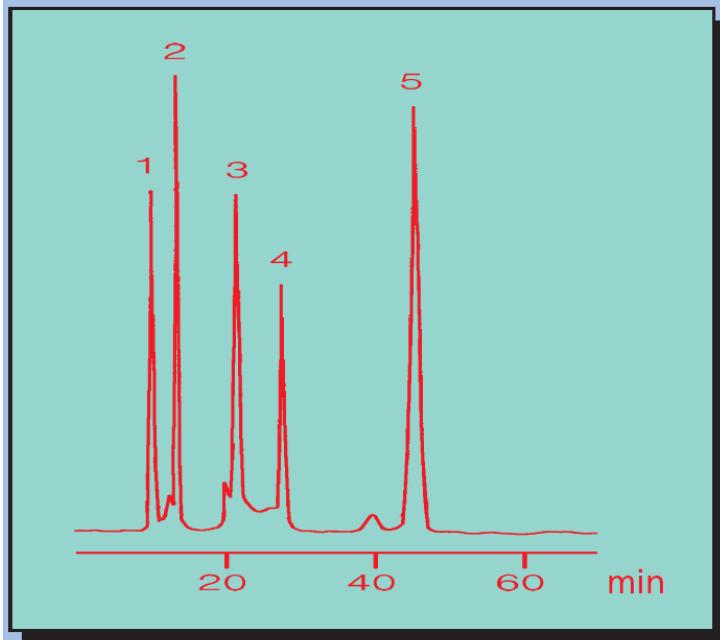


# Traditional Ion Exchange Chromatography

Salt gradient from low salt to high salt concentration

09 105

## Separation of Proteins by Ion exchange HPLC



- 1) Trypsinogen
- 2) Ribonuclease A
- 3) Cytochrome C
- 4) Chymotrypsinogen
- 5) Lysozyme

Column phase: GROM-SIL 300 WCX, 7 µm

Column size: 250 x 4.6 mm

Eluent A: 0.05 M  $\text{NaH}_2\text{PO}_4$ , pH 6.4  
B: 0.5 M  $\text{NaH}_2\text{PO}_4$ , pH 6.4

Gradient: 0-18% B (0-21 min), 18% B (21-26 min),  
18-60% B (26-70 min)

Flow rate: 1.0 ml/min

Pressure: 7 MPa

Temperature: RT

Detection (UV): 280 nm

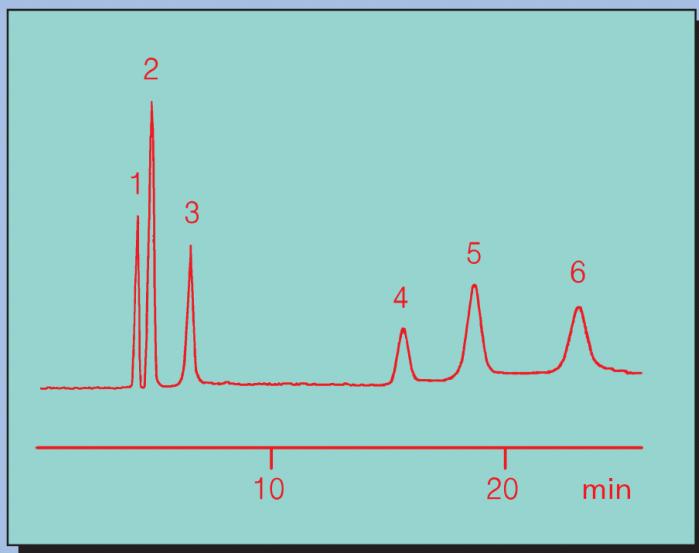
Injection: 100 µl

# Modern Ion Exchange Chromatography

3 possibilities for elution:

- Salt gradient from low salt to high salt concentration\*
- pH shift / gradient
- ACN or MeOH gradient

## 07 127 Separation of Nucleotides



- 1) GMP
- 2) AMP
- 3) IMP
- 4) GDP
- 5) ADP
- 6) IDP

Column phase: GROM-SIL 120 Amino-3 CP, 5  $\mu$ m  
Column size: 250 x 4.6 mm  
Eluent A: 5 mM  $\text{KH}_2\text{PO}_4$ , pH 2.9, 15% ACN  
B: 500 mM  $\text{KH}_2\text{PO}_4$ , pH 4.0, 15% ACN  
Gradient: 40 - 100% B (0-20 min), 100% B (20-25 min)  
Flow rate: 2 ml/min  
Pressure: 14 MPa  
Temperature: RT  
Detection (UV): 254 nm  
Injection: 10  $\mu$ l (160  $\mu$ g/ml)

\* For MS applications use volatile buffers up to 100 mM

# Sterical - Multi Selectivity Effects

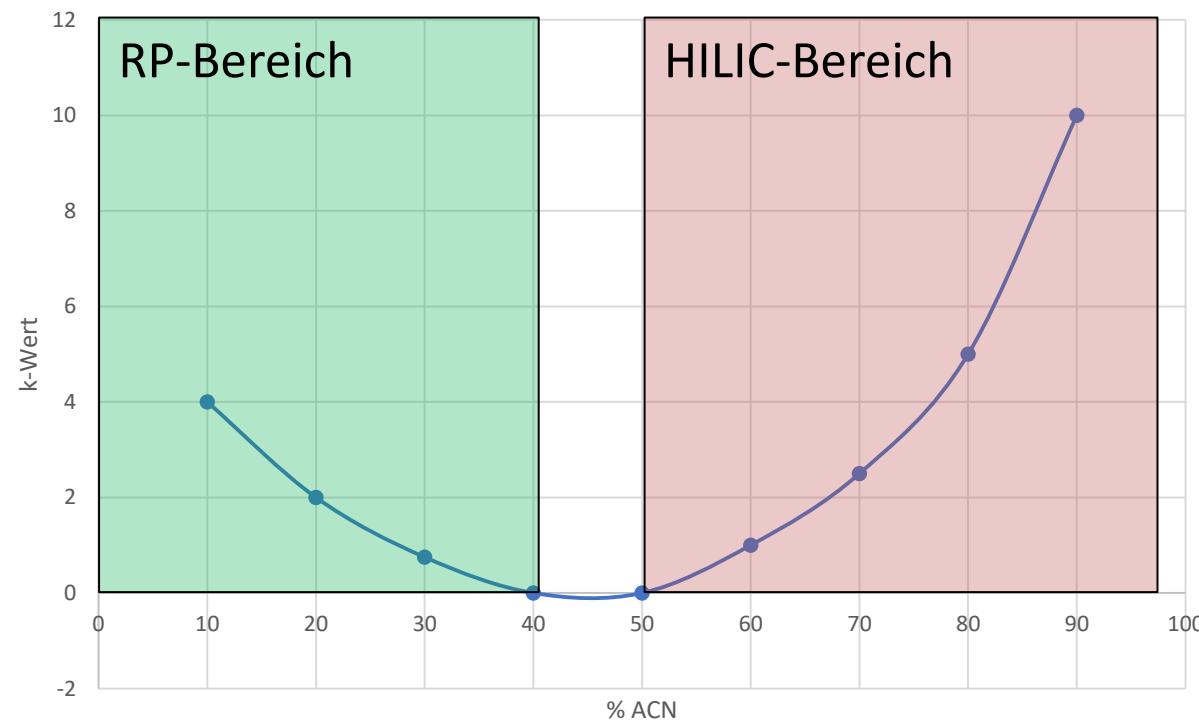
- Most of the RP/IEX phases are bonded with the RP and the IEX group: 2 different silanes are used
- The modern Mixed Mode Phases have the RP and the IEX group as one silane: the RP and IEX group has always the same distance and the sample molecule has two interaction points with a defined sterically orientation – goes a step ahead to Affinity Chromatography
- The similar situation we have with the Phenyl Phases:
  - Propy-Phenyl: no mobility of the Phenyl group
  - Hexyl-Phenyl: high mobility of the Phenyl group
  - Biphenyl: two Phenyl groups in a chain with a defined orientation and distance
  - Diphenyl: two Phenyl groups in tetraeder angle with a defined orientation and distance

# Mobile Phase Strategies for RP / IEX

- Starting at low salt and low organic modifier:  
gives maximum retention for RP and IEX groups
- Keeping the salt concentration low and increase org. modifier  
IEX keeps longer active and RP elution starts
- Increasing salt concentration and org. modifier  
IEX and RP elution starts
- Using MeOH as org. modifier  
supports IEX / dissociation and leads to more retention
- Using ACN as org. modifier  
reduces IEX / dissociation and therefore retention

# Selectivity in HPLC – charge situations

Amino Phase: Aminopropyl

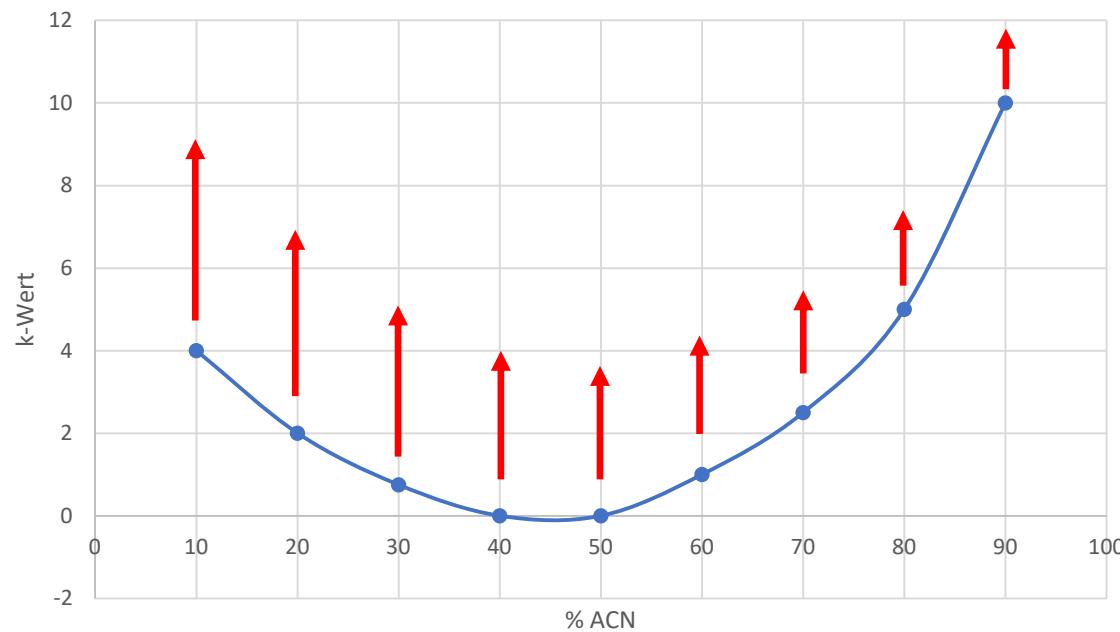


# Selectivity in HPLC – charge situations

Amino Phase:

Aminopropyl at a pH of 3-7 in the mobile Phase

- Acidic Molecules: Increasing Retention – additional IEX
- More Acetonitrile – lower dissociation

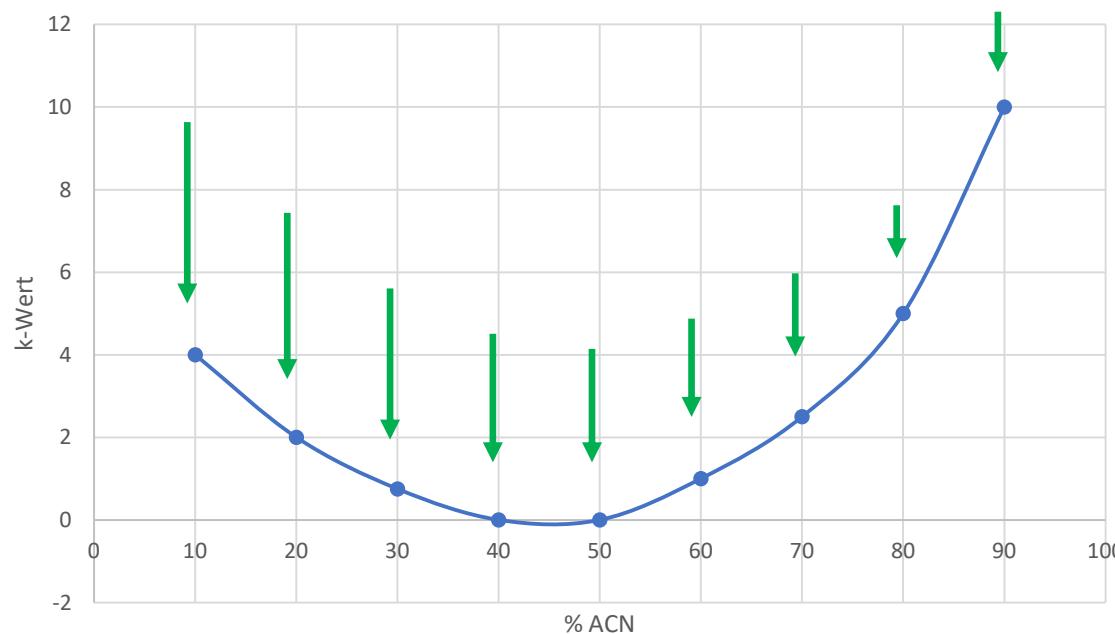


# Selectivity in HPLC – charge situations

Amino Phase:

Aminopropyl bei pH-Wert im Eluent 3-7

- basic Molecules: decreasing retention
- more Acetonitrile – lower Dissociation



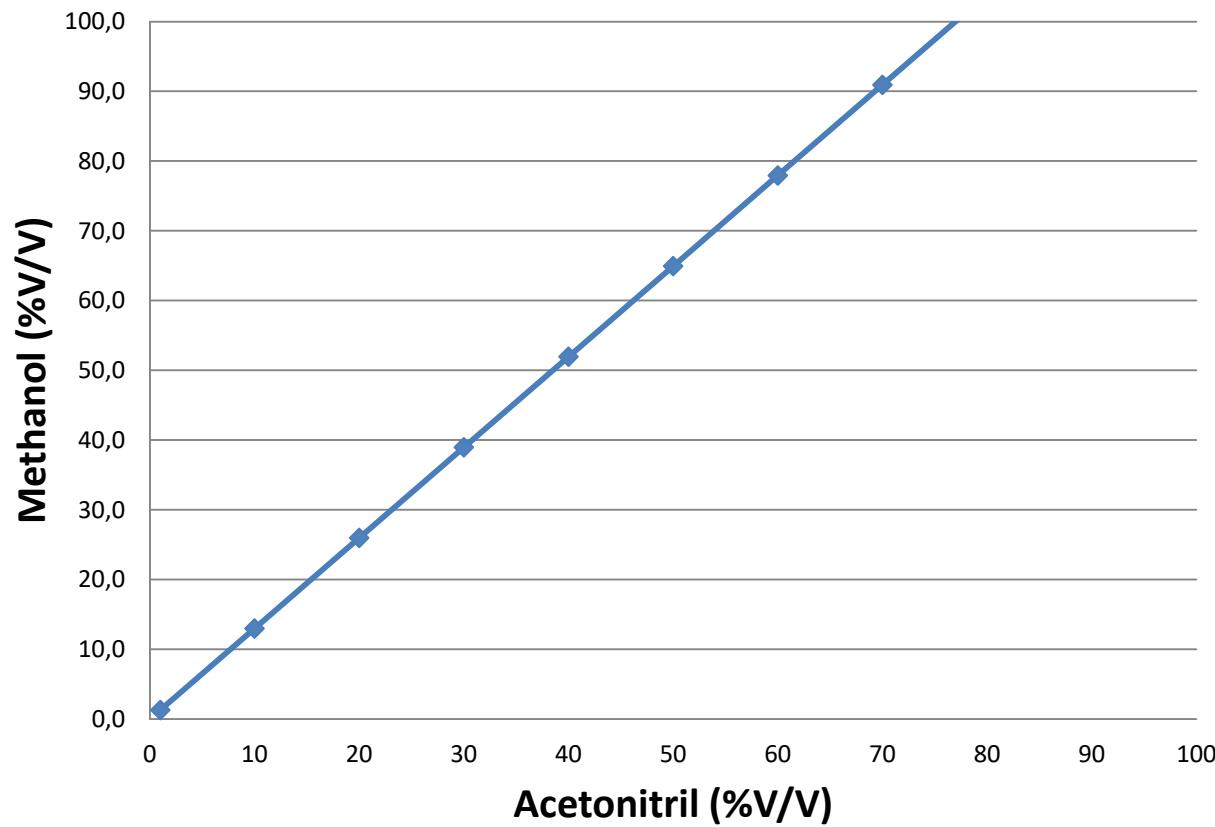
# Influence of Mobile Phase

Conversion of the eluent composition  
with the solubility parameter  $\delta$

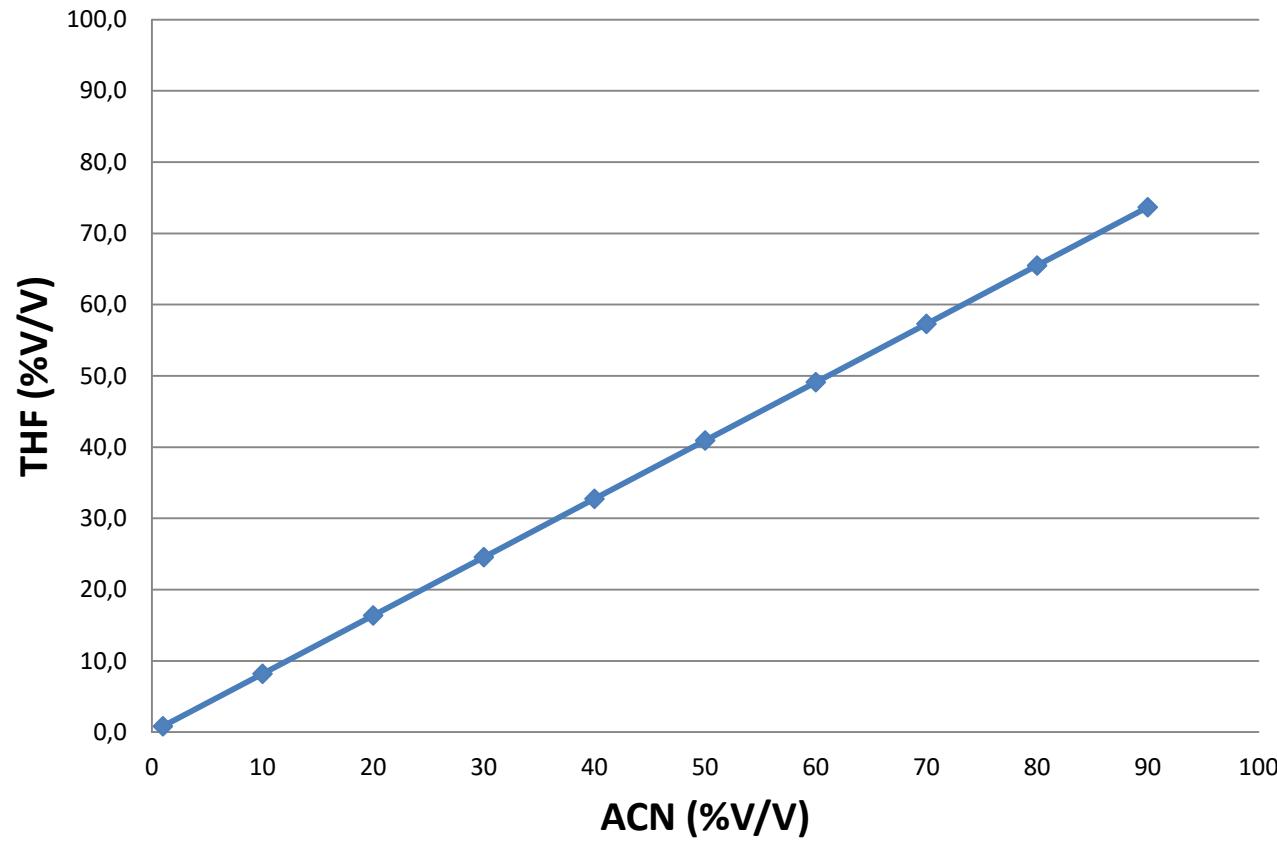
Acetonitril	$\delta_A$	23,9	$\delta =$	Solubility parameter
Water	$\delta_W$	47,8	$\phi =$	Volume part
Methanol	$\delta_M$	29,4		
Mixture	$\delta_m$	35,85		

$$\delta_m = \sum_i \phi_i \delta_i$$

# Influence of Mobile Phase



# Influence of Mobile Phase



# Influence of Mobile Phase

Column test mix:

Uracil

Phenol

N,N-Diethyl-M-Toluamide

Toluone

Standard eluent:

ACN : Water = 58 : 42

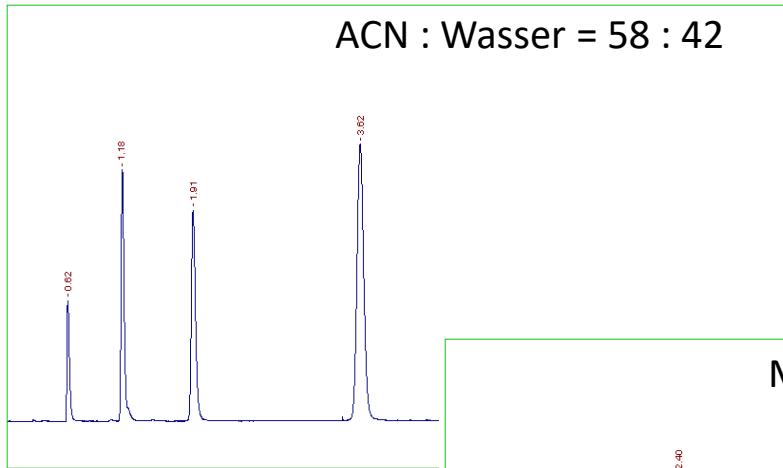
calculated:

MeOH : Water = 75 : 25

adjusted:

MeOH : Water = 65 : 35

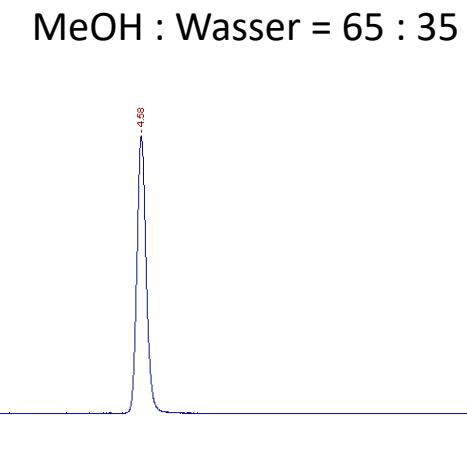
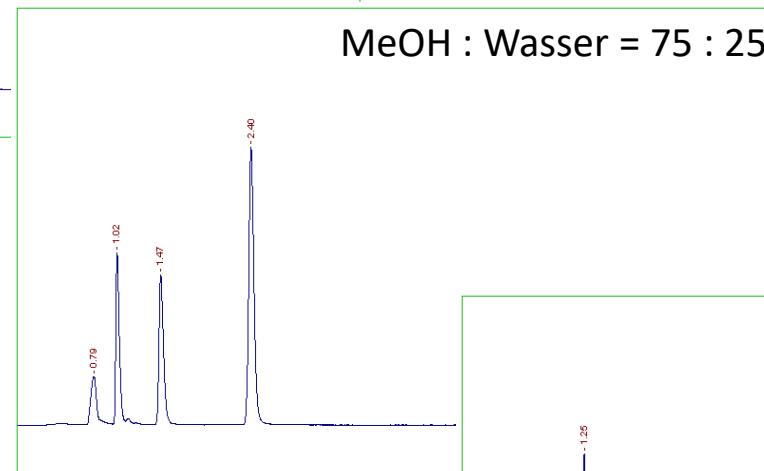
# Influence of Mobile Phase



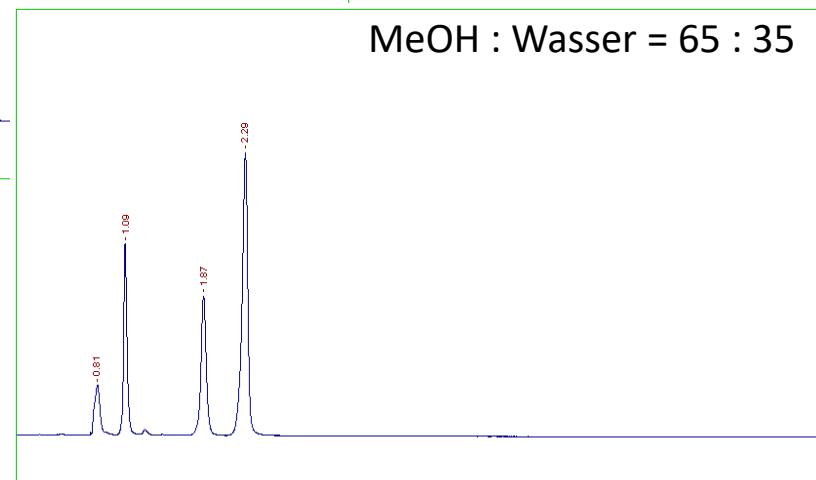
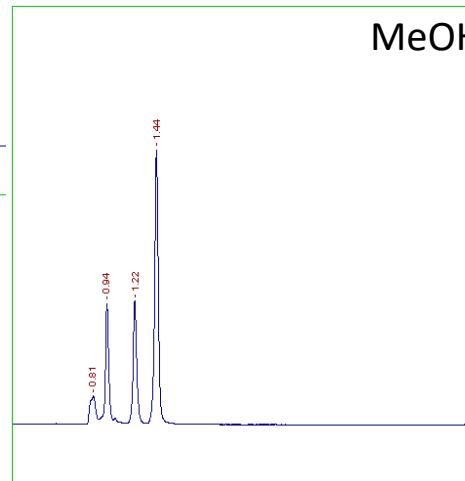
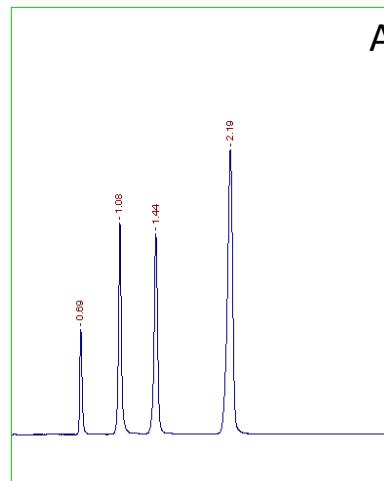
Standard C18

Reprospher C18, 3 µm;  
100 x 2 mm

Standard C18



# Influence of Mobile Phase



Polar C18

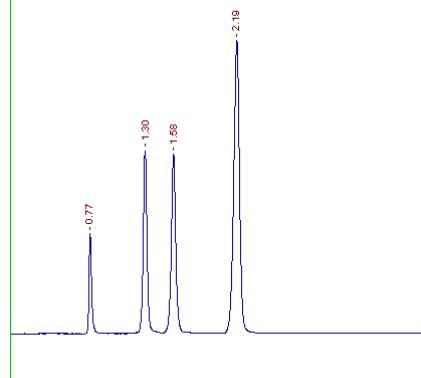
Platinum C18, 3 µm;  
100 x 2 mm

Polare C18 Phase

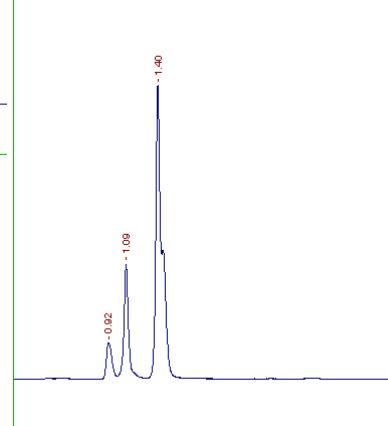
# Influence of Mobile Phase

Phenyl - Phase

ACN : Wasser = 58 : 42



MeOH : Wasser = 75 : 25



MeOH : Wasser = 65 : 35

